

DISCREPANCIES BETWEEN CEREBRAL COMPUTED TOMOGRAPHY AND WESTERN BLOT IN THE DIAGNOSIS OF NEUROCYSTICERCOSIS

HECTOR H. GARCIA, GENARO HERRERA, ROBERT H. GILMAN, VICTOR C. W. TSANG, JOY B. PILCHER, JOSE F. DIAZ, ERROL J. CANDY, ELBA MIRANDA, AND JORGE NARANJO, AND THE CYSTICERCOSIS WORKING GROUP IN PERU (CLINICAL STUDIES COORDINATION BOARD)*

Universidad Peruana Cayetano Heredia, Lima, Peru; Department of International Health, Johns Hopkins University School of Hygiene and Public Health, Baltimore, Maryland; Parasitic Diseases Branch, Division of Parasitic Diseases, National Center for Infectious Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia; Department of Neuroradiology, Johns Hopkins Hospital, Baltimore, Maryland

Abstract. Serum samples from sequential patients who underwent cerebral computed axial tomography (CT) scan in a Peruvian radiologic clinic were tested by the highly sensitive and specific enzyme-linked immunoelectrotransfer blot (EITB) test to detect antibodies to *Taenia solium*. The results of the EITB test were compared with those obtained by CT scan for the diagnosis of neurocysticercosis. Of the 383 patients sampled, 32 (8%) were seropositive. The results of CT and EITB were frequently discrepant. When compared with the EITB assay, the CT scan was 44% sensitive and 95% specific. The sensitivity of CT increased to 63% if less specific images (single calcifications, granulomas, or hydrocephalus) were included. The CT scan for diagnosis of cysticercosis can best be used in conjunction with a reliable serologic test such as the EITB.

Taenia solium cysticercosis is an important parasitic disease affecting the human central nervous system (CNS).^{1,2} Cysticercosis is common in those developing countries where free-ranging pigs are raised.³ The life cycle of this cestode requires an intermediate host, normally the pig, for the cystic form of the parasite and a definitive host, humans, for the adult tapeworm.⁴ However, humans infected by *Taenia* eggs serve as the intermediate host and develop the cystic form. Besides the CNS, cysts may also be found in the subcutaneous tissue, muscle, and eye,^{5,6} and more rarely in other parts of the body.⁷

Many factors affect the clinical presentation of cysticercosis, including the number, size, lo-

cation, and form of the cysts and the immune response of the host. The interaction of these variables results in a wide spectrum of neurologic symptoms.^{1,2,5,7}

Cerebral computed axial tomography (CT) has been the method of choice for the diagnosis of neurocysticercosis.^{1,5} However, CT images are not pathognomonic for this disease and apparently positive test findings can be caused by a variety of other conditions.⁸⁻¹⁰ The CT scan has been claimed to have a sensitivity and specificity of about 95%,^{1,11} but controlled studies are not available. Approximately 25% of cysticercosis cases occur in the racemose form, in which the cysts are located at the base of the brain. Such basal cysts appear on CT only as hydrocephalus or deformation of the basal cisterns.^{4,12} Cysticerci also present as granulomas or nodular forms, which often are indistinguishable from other granulomatous conditions such as tuberculosis.^{8,13,14}

Evaluation of the sensitivity and specificity of the CT scan for the diagnosis of cysticercosis requires a standard for comparison. Until recently, the only standards available for testing the accuracy of CT were the results obtained by postmortem examination or neurosurgical exploration.¹⁵ The recently developed enzyme-linked

* The other members of the Cysticercosis Working Group in Peru are P. Torres, C. Gallo, C. Carcamo, M. Verastegui, T. Montenegro, M. Alvarez (Universidad Peruana Cayetano Heredia); C. Evans (Cambridge University, UK); A. E. Gonzales (Universidad Nacional Mayor de San Marcos); M. Castro, A. Guerrero (A B PRISMA); M. Martinez, M. Porras, M. Alvarado, E. Orrillo, L. Palomino, G. Alban, L. Calagua, S. Escalante, L. Trelles, O. Aliaga, N. Rios-Saavedra, M. Velarde, J. M. Cuba, H. Estrada, M. Soto, L. Portilla (Instituto Nacional de Ciencias Neurologicas); A. Terashima, J. Cabrera, P. Campos (Hospital Cayetano Heredia); D. Morote (Hospital Edgardo Rebagliati); U. Rocca (Hospital Guillermo Almenara).

immuno-electrotransfer (EITB) assay^{16,17} provided a sensitive and specific method for diagnosing *T. solium* infection. We used the EITB assay to compare the sensitivity and specificity of CT scans in patients admitted to a radiologic clinic for the diagnosis of neurologic

MATERIALS

The study was conducted in a radiologic clinic center in Lima, Peru. A list of patients was obtained, blood samples were collected for EITB from patients during their working hours for cerebral computed axial tomography (CT) during two separate periods: January 12, 1988 and February 1988. Sampling was often not possible during working hours and episodes of emergency working hours for the clinic were between 8:00 AM and 1:00 PM, and occasionally, patients were present during normal working hours. These emergency patients, in addition, on a few occasions, were unable to visit the clinic. A sample was taken during the study. The study was approved by the ethics committees of both the Universidad Cayetano Heredia and the Johns Hopkins

Axial computed tomography

All scans were performed using a Siemens scanner (Elscent, Haifa, Israel). A 5 mm scan circle and a 5 mm slice thickness image series were made. All patients were examined using a contrast injection (iodamide contrast medium, Iodamide 370, 1 ml/kg; Schering, Kenilworth, NJ). The scans were read by a radiologist who did not know the clinical history of the patient. The CT scans of patients who were known to have cysticercosis were read blind to the radiologist.

Axial computed tomography for the diagnosis of cysticercosis

Radiologic criteria for the diagnosis of neurocysticercosis by CT are as follows: used the following criteria

immunoelectrotransfer blot (EITB, Western Blot) assay^{16,17} provides a precise method of diagnosing *T. solium* infection. In this study, we used the EITB assay as a standard to determine the sensitivity and specificity of cerebral CT in patients admitted to a radiologic center for diagnosis of neurologic symptoms.

MATERIALS AND METHODS

The study was conducted in a private radiology center in Lima, Peru. After informed consent was obtained, blood samples were collected for EITB from patients admitted during normal working hours for cerebral CT examination during two separate periods (September 16–October 12, 1988 and February 6–May 2, 1989). Sampling was often not possible during clinic off hours and episodes of labor unrest. Normal working hours for the clinic are between 9:00 AM and 1:00 PM, and from 4:00 PM to 8:00 PM. Occasionally, patients are admitted outside of normal working hours on an emergency basis. These emergency patients are not sampled. In addition, on a few occasions, our study nurse was unable to visit the clinic; therefore, no sample was taken during these rare occasions. The study was approved by the ethical review boards of both the Universidad Peruana Cayetano Heredia and the Johns Hopkins University.

Axial computed tomography scans

All scans were performed on an Exel 2002 scanner (Elscent, Haifa, Israel) with a 140–480-mm scan circle and a 5.3–5.8 sec scan time. Two image series were made using 10-mm slices. Patients were examined before and after contrast injection (iodamide metylglucamate, Uromiron[®], 1 ml/kg; Schering G.A., Berlin, Germany). The scans were read by a neuroradiologist (GH) who did not know the results of the EITB assay. The CT scans of patients who were diagnosed as having cysticercosis but had a negative EITB result were read blindly by a second neuroradiologist.

Axial computed tomography scan diagnosis of cysticercosis

Radiologic criteria for the diagnosis of neurocysticercosis by CT are not uniform.^{11,18,19} We used the following criteria, modified from a pre-

vious study:²⁰ 1) one or more cystic images and/or 2) two or more compatible calcifications. Calcifications were considered compatible if they were not located in zones of physiological calcifications,²¹ were annular or rounded, and were less than 10 mm in diameter.

Other diagnosis

The neuroradiologist's reports were classified as follows: 1) normal CT scan, 2) cysticercosis, 3) vascular disease, and 4) neoplasia. We also noted the presence of 1) cystic images, 2) abnormal calcifications, 3) granulomatous images, 4) hydrocephalus, and 5) cortical or subcortical atrophy.

Enzyme-linked immunoelectrotransfer blot assay

The EITB assay for *T. solium*-specific antibodies was performed as previously described.¹⁶ Briefly, seven lentil-lectin purified *T. solium* glycoprotein antigens are used in an immunoblot format to detect infection-specific antibodies in serum or cerebrospinal fluid (CSF) samples. The assay was performed in the Accutran system (Schleicher & Schuell, Keene, NH). Antibody reactions against these glycoproteins in serum and/or CSF were visualized with the H₂O₂/diaminobenzidine (DAB) substrate system.²² An antibody reaction to one or more glycoprotein bands was designated a positive result. Equivocal assays were repeated. Assays were performed at the Laboratory of Parasitology of the Universidad Peruana Cayetano Heredia, and duplicate specimens were sent to the Centers for Disease Control and Prevention (CDC, Atlanta, GA) for quality control.

RESULTS

Sample characteristics

Three hundred eighty-three (63%) of 606 patients classified underwent cerebral CT had a blood sample taken for the EITB. Comparison of the sampled patients with those not sampled showed that the two groups were similar in age, sex ratio, and the proportion of patients diagnosed as having cysticercosis or neoplasia (Table 1). However, the sampled group had significantly ($P < 0.001$) more normal CT scans, probably because the majority of these patients

ED NOSIS

NDY, ING GROUP

Health, Johns
asitic Diseases
Centers for
Johns Hopkins

bral computed
by the highly
to detect an-
those obtained
pled, 32 (8%)
then compared
sensitivity of
nas, or hydro-
est be used in

s and the immune
interaction of these
spectrum of neuro-

tomography (CT)
e for the diagnosis
ever, CT images
his disease and ap-
s can be caused by
10 The CT scan has
ivity and specificity
led studies are not
% of cysticercosis
form, in which the
of the brain. Such
y as hydrocephalus
cisterns.^{4,12} Cysti-
olomas or nodular
istinguishable from
ions such as tuber-

ty and specificity of
sis of cysticercosis
comparison. Until re-
available for testing
results obtained by
neurosurgical explo-
oped enzyme-linked

TABLE 1

Comparison of sampled and unsampled patients undergoing cerebral computed tomography (CT) scan examination

	Sampled (n = 383)	Unsampled (n = 223)	P*
Age, years (mean \pm SD)	40 \pm 22	39 \pm 28	NS
No. (%) males	223 (58)	122 (55)	NS
CT diagnosis, no. (%)†			
Normal CT scan	114 (30)	31 (14)	<0.001
Trauma	32 (8)	41 (18)	<0.001
Vascular disease	86 (22)	74 (33)	<0.01
Neoplasia	35 (9)	17 (8)	NS
Cysticercosis	30 (8)	12 (5)	NS
Others	110 (29)	74 (33)	NS
CT characteristics, no. (%)			
Hydrocephalus	30 (8)	28 (13)	NS
Atrophy	72 (19)	47 (21)	NS
Calcifications	38 (10)	13 (6)	NS

* P was determined by the chi-square test. The difference in mean age was tested by student's *t*-test. NS = not significant.

† Some patients had more than one pathology in the same CT scan.

came during working hours. In contrast, patients with acute trauma or vascular accidents often were not sampled because they came to the clinic at irregular hours when sampling did not occur (Table 1).

Results of EITB assay

Of the 383 patients sampled, 32 (8%) had EITB assay results positive for antibodies to *T. solium*. No differences in age or sex distribution were found in EITB-positive versus EITB-negative patients (Table 2).

Results of the EITB and CT

Thirty patients were diagnosed as having cysticercosis by EITB and 32 by CT scan. The results obtained in the CT scan and the EITB assay were frequently discrepant. More than 50% of the cases diagnosed as cysticercosis by the radiologist were EITB-negative (Table 2). Compared with the EITB, the CT scan was only 44% sensitive, detecting 14 of 32 EITB-positive patients. However, CT and EITB diagnosis of cysticercosis were significantly associated when the whole study population is examined. No patient with CT-diagnosed neoplasia had positive EITB

TABLE 2

Characteristics of enzyme-linked immunoelectrotransfer blot (EITB)-positive and EITB-negative patients and comparison of results with computed tomography (CT) scan diagnosis

	EITB+ (n = 32)	EITB- (n = 351)	P*
Age, years (mean \pm SD)	45 \pm 20	40 \pm 22	NS
Age, years (range)	9-82	0-85	
No. (%) males	20 (63)	202 (58)	NS
CT diagnosis, no. (%)†			
Normal CT scan	5 (16)	109 (31)	NS
Trauma	2 (6)	30 (9)	NS
Vascular disease	5 (16)	81 (23)	NS
Neoplasia	0 (0)	35 (10)	NS
Cysticercosis	14 (44)	16 (5)	<0.001
Others†	8 (25)	102 (29)	NS

* P was determined by either Fisher's exact test or chi-square test. The difference in mean age was tested by Student's *t*-test. NS = not significant.

† Some patients had more than one pathology in the same CT scan.

results, and patients have a lower percentage (Table 2). The overall sensitivity was 44% from dividing all CT scan results by the results of all EITB-negative patients (95%).

The 18 patients with positive CT scan results and negative EITB assay had a variety of pathologic characteristics. Four had atrophy, three had hydrocephalus, two had hydrocephalus and atrophy, one had atrophy and one had a granuloma.

There were 16 patients with negative EITB but positive CT scan. On the original CT report, six of these were inactive cysticercosis, the remaining 10 had other pathologies (active cysticercosis with calcifications, trauma, vascular disease). These were reread by another radiologist who also blinded to the EITB results. The 10 not having cysticercosis were eight as having inactive cysticercosis and two of the 10 patients with active cysticercosis by the EITB were active by the second reading.

Subsequent to the EITB assay, 16 patients had at least one CT scan examined by EITB, in 16 samples. All samples were

In this study, we used a specific EITB assay for antibodies to *T. solium* in comparison with the results of the CT scan for the diagnosis of cysticercosis. Eight percent of the patients with a positive EITB test result had a CT scan result that cysticercosis is

When measured by the sensitivity of the CT scan for specific images such as multiple nodules and single calcifications, the overall sensitivity was 44%. We can only determine the sensitivity of the EITB assay with *T. solium* but not the sensitivity of the CT scan located in the CNS. The sensitivity of the EITB assay compared between a negative EITB diagnosis is not necessarily but may represent the

phy (CT) scan exami-

P*
NS
NS
<0.001
<0.001
<0.01
NS
NS
NS
NS
NS
NS

ificant.

diagnosed as having cysticercosis by CT scan. The results of the EITB assay and the EITB assay. More than 50% of patients diagnosed as having cysticercosis by the radiologist. Computed tomography scan was only 44% sensitive. EITB-positive patients were 52%. EITB diagnosis of cysticercosis was associated when the patient was examined. No patient was found to have had positive EITB

EITB-negative patients and

P*
NS
NS
NS
NS
NS
NS
<0.001
NS

*Fisher's exact test. NS = not significant.

results, and patients with normal scans tended to have a lower percent of EITB seropositivity (Table 2). The overall specificity for CT, derived from dividing all CT-negative patients (335) by all EITB-negative patients (351), was high (95%).

The 18 patients with positive EITB but negative CT scan results had the following tomographic characteristics: five appeared normal, four had atrophy, three had a single calcification, two had hydrocephalus, two had subdural hematomas, one had a surgical sequel for trauma, and one had a granuloma.

There were 16 patients whose results were EITB negative but CT scan positive. In the initial CT report, six of them were diagnosed with inactive cysticercosis (only calcifications), and the remaining 10 had at least one cystic image (active cysticercosis), in five cases coexisting with calcifications. Fourteen of those 16 scans were reread by another neuroradiologist (EJC), also blinded to the EITB. Five were reported as not having cysticercosis (three as normal) and eight as having inactive cysticercosis. Only one of the 10 patients diagnosed as having active cysticercosis by the first reader was considered active by the second (Table 3).

Subsequent to this study, eight of these 16 patients had at least one additional sample examined by EITB, including six with CSF samples. All samples were negative.

DISCUSSION

In this study, we used the highly sensitive and specific EITB assay for the detection of antibodies to *T. solium* infection to assess the accuracy of the CT scan for the diagnosis of cysticercosis. Eight percent of the patients scanned had a positive EITB test result, providing further evidence that cysticercosis is endemic in Peru.^{20, 23}

When measured by the EITB test, the sensitivity of the CT scan was only 44%. If less specific images such as hydrocephalus, granulomas, and single calcifications were considered, the overall sensitivity was 63%. Because the EITB can only determine whether a patient is infected with *T. solium* but not whether the infection is located in the CNS, the EITB will underestimate the sensitivity of the CT scan. Thus, discordance between a negative CT and a positive serodiagnosis is not necessarily due to misdiagnosis but may represent the presence of an adult worm

TABLE 3

Reproducibility of neuroradiologic diagnosis in a group of 14 computed tomography-positive enzyme-linked immunoelectrotransfer blot (EITB)-negative patients

	Active cysticercosis	Inactive cysticercosis	No cysticercosis
Neuroradiologist 1	10	4	—*
Neuroradiologist 2	1	8	5

* Reports on 14 patients who were diagnosed as having cysticercosis by the first radiologist and who had a negative EITB result. Scans of two patients were not recovered from the archives.

or extracerebral cysts. Another possibility is that some patients have the racemose form of the disease, in which cysts are located at the base of the brain. Basally located cysts often do not show up on a CT scan except as hydrocephalus.¹ Magnetic resonance imaging (MRI) may detect these lesions as well as small intraparenchymal lesions missed on a CT scan;^{24, 25} however, MRI scanners are rarely available in endemic areas.

Certainly in patients with multiple calcifications, which are end-stage lesions, the active disease that caused the calcifications cannot be known. Moreover, EITB-negative patients with calcified scars or hydrocephalus may have had active cysticercosis in the past but their immunity may have waned. It is still not known how long antibody persists in patients whose cysts have resolved. Regrettably, the patients in this study were referrals to a private radiology center for CT examinations only. Results of the EITB and CT were reported to their respective primary physicians, who provided appropriate treatment. Within the constraints of the present study, therefore, we were not able to provide direct treatment or follow-up for these patients.

The specificity of CT is high (95%) when all patients are included. When positive CT results were compared with EITB assay results, the specificity of CT was much poorer. More than 50% of the patients diagnosed as having cysticercosis by CT were EITB negative. There are several possible reasons for this. First, although the specificity of the EITB test has not been questioned, its sensitivity may vary with specific lesions.^{16, 20} A recent report states that single intraparenchymal lesions are often associated with a negative EITB test result.²⁰ Unfortunately, the population of this study was self-selected, and thus true sensitivity cannot be calculated. Second, different readers may have a wide variance

in CT interpretation,²⁷ especially for small lesions. Finally, overdiagnosis probably occurs because of multiple other causes, including fungal infections, chronic abscesses, tuberculosis, and cystic neoplasms, for images seen on scans reported as cysticercosis.

In an endemic zone, cerebral CT was not read as cysticercosis in more than one-third of the EITB-positive neurologic patients. The diagnosis of this disease will be markedly improved if CT is used in conjunction with an accurate immunologic assay such as the EITB.

Acknowledgments: We thank the Tomografía Axial Computarizada e Imagenología, Sociedad Anónima, Radiological Center for cooperation, Reyda Terrones for technical assistance, Dr. R. Berendson for advice, and A. B. Prisma for aid in data analysis.

Financial support: This study was supported by grants from the International Development Research Center, AID-PSTC-7208, and the Robert Gilman ER Fund.

Disclaimer: The use of trade names is for identification only and does not imply endorsement by the Public Health Service or by the U.S. Department of Health and Human Services.

Authors' addresses: Hector H. Garcia, Genaro Herrera, Jose F. Diaz, Elba Miranda, and Jorge Naranjo, Universidad Peruana Cayetano Heredia, Lima, Peru. Robert H. Gilman, Department of International Health, Johns Hopkins University School of Hygiene and Public Health, Baltimore, MD 21205. Victor C. W. Tsang and Joy B. Pilcher, Parasitic Diseases Branch, Division of Parasitic Diseases, National Center for Infectious Diseases, Centers for Disease Control and Prevention, Atlanta, GA 30333. Errol J. Candy, Department of Neuroradiology, Johns Hopkins Hospital, Baltimore, MD 21205.

REFERENCES

- Nash TE, Neva FA, 1984. Recent advances in the diagnosis and treatment of cerebral cysticercosis. *N Engl J Med* 311: 1492-1496.
- Grisolia JS, Wiederholt WC, 1982. CNS cysticercosis. *Arch Neurol* 39: 540-544.
- Mahajan RC, 1982. Geographical distribution of human cysticercosis. Flisser A, Williams K, Lacleite JP, Larralde C, Ridaura C, Beltran F, eds. *Cysticercosis: Present State of Knowledge and Perspectives*. New York: Academic Press, 39-46.
- Malagon F, 1989. Elementos del binomio taeniasis/cisticercosis. Una síntesis. Flisser A, Malagon F, eds. *Cisticercosis Humana y Porcina*. Mexico D.F.: Limusa, 3-6.
- Loo L, Braude A, 1982. Cerebral cysticercosis in San Diego: a report of 23 cases and a review of the literature. *Medicine (Baltimore)* 61: 341-359.
- Lozano D, 1983. Ophthalmic cysticercosis. Palacios E, Rodriguez-Carbajal J, Taveras J, eds. *Cysticercosis of the Central Nervous System*. Springfield, IL: Charles C. Thomas, 84-100.
- McCormick GF, Zee CS, Heiden J, 1982. Cysticercosis cerebri. Review of 127 cases. *Arch Neurol* 39: 534-539.
- Enzmann DR, Norman D, Mani MD, Newton TH, 1976. Computed tomography of granulomatous basal arachnoiditis. *Radiology* 120: 341-344.
- Whelan MA, Stern J, 1981. Intracranial tuberculoma. *Radiology* 138: 75-81.
- Zee CS, Segall HD, Miller C, Tsai FY, Teal JS, Hieshima G, Ahmadi J, Halls J, 1980. Unusual neuroradiological features of intracranial cysticercosis. *Radiology* 137: 397-407.
- Rodriguez-Carbajal J, Palacios E, Chi-Shing Z, 1983. Neuroradiology of cysticercosis of the central nervous system. Palacios E, Rodriguez-Carbajal J, Taveras J, eds. *Cysticercosis of the Central Nervous System*. Springfield, IL: Charles C. Thomas, 101-143.
- Rabiela-Cervantes M, Rivas-Hernandez A, Castillo-Medina S, Gonzales-Angulo A, 1985. Pruebas morfológicas de que *C. cellulosae* y *C. racemosus* son larvas de *Taenia solium*. *Arch Invest Med (Mex)* 16: 83-92.
- Witham R, Johnson R, Roberts D, 1979. Diagnosis of miliary tuberculosis by cerebral computerized tomography. *Arch Intern Med* 139: 479-480.
- Nava J, 1983. La cisticercosis del sistema nervioso central. *Salud Publica Mex* 25: 297-300.
- Flisser A, Madrazo I, Gonzales D, Sandoval M, Rodriguez-Carbajal J, De Dios J, 1988. Comparative analysis of human and porcine cysticercosis by computed tomography. *Trans R Soc Trop Med Hyg* 82: 739-742.
- Tsang VCW, Brand J, Boyer E, 1989. Enzyme-linked immunoelectrotransferency blot assay and glycoprotein antigens for diagnosing human cysticercosis (*Taenia solium*). *J Infect Dis* 159: 50-59.
- Diaz JF, Verastegui M, Gilman RH, Tsang VCW, Pilcher JB, Gallo C, Garcia HH, Torres P, Montenegro T, Miranda E, and the Cysticercosis Working Group in Peru (CWG), 1992. Immunodiagnosis of human cysticercosis (*Taenia solium*): a field comparison of an antibody-enzyme-linked immunosorbent assay (ELISA), an antigen-ELISA, and an enzyme-linked immunoelectrotransfer blot (EITB) assay in Peru. *Am J Trop Med Hyg* 46: 610-615.
- Mervis B, Lotz JW, 1980. Computed tomography (CT) in parenchymatous cerebral cysticercosis. *Clin Radiol* 31: 521-528.
- Byrd S, Locke G, Biggers S, Percy A, 1982. The computed tomography appearance of cerebral cysticercosis in adults and children. *Radiology* 144: 819-823.
- Garcia HH, Martinez M, Gilman RH, Herrera G, Tsang VCW, Pilcher JB, Diaz F, Verastegui M, Gallo C, Porras M, Alvarado M, Naranjo J, Miranda E, 1991. Di-
 21. Cardenas Cardenas las calcificaciones nostico neuroquir 249-267.
 22. Tsang VCW, Peralt zyme-linked imm niques (EITB) fo antigens and anti trophoresis. *Meth*
 23. Garcia HH, Gilma VCW, Pilcher JB, M, Miranda E, 19 cause of epilepsy i
 24. Barkovich A, Citrin

- cysticercosis. Palatal J, Taveras J, eds. *Cerebral Nervous System*. Thomas, 84-100.
- Wideman J, 1982. Cysticercosis of 127 cases. *Arch Neurol* 39: 131-134.
- Wang MD, Newton TH, et al. Pathology of granulomatous disease. *Neuropathology* 120: 341-344.
- Intracranial tuberculoma. *Neuropathology* 120: 345-347.
- Chen C, Tsai FY, Teal JS, et al. Unusual case of intracranial cysticercosis. *Arch Neurol* 197-407.
- Wang E, Chi-Shing Z, et al. Cysticercosis of the brain. *Cysticercosis of the Brain*. Springfield, IL: Charles C. Thomas, 1943.
- Hernandez A, Castilligulo A, 1985. Pruebas de inmunodiagnóstico de *C. cellulosae* y *C. raiaenia solium*. *Arch Neurol* 42: 123-125.
- Wang D, 1979. Diagnosis of cerebral cysticercosis. *Arch Intern Med* 139: 123-125.
- Wang del sistema nervioso. *Arch Mex* 25: 297-300.
- Wang D, Sandoval M, Dios J, 1988. Common and porcine cysticercosis. *Trans R Soc Trop Med Hyg* 62: 123-125.
- Wang E, 1989. Enzyme-linked immunoblot assay and diagnosis of human cysticercosis. *J Infect Dis* 159: 123-125.
- Wang RH, Tsang VCW, et al. (HH, Torres P, Monard) the Cysticercosis (WG), 1992. Immunodiagnosis of *Taenia solium* cysticercosis (Taenia solium) of an antibody-dependent assay (ELISA), an enzyme-linked immunoblot assay in Peru. *Am J Trop Med Hyg* 46: 15.
- Computed tomography of cerebral cysticercosis. *Arch Neurol* 47: 520-523.
- Percy A, 1982. The clearance of cerebral cysticercosis in children. *Radiology* 143: 123-125.
- Wang RH, Herrera G, et al. (az F, Verastegui M, Naranjo J, Miranda E, 1991. Diagnosis of cysticercosis in endemic zones. *Lancet* 338: 549-551.
21. Cardenas Cardenas J, 1950. Valor radiológico de las calcificaciones intracraneales para el diagnóstico neuroquirúrgico. *Gac Med Mex* 80: 249-267.
22. Tsang VCW, Peralta JM, Simons AR, 1983. Enzyme-linked immunoelectrotransfer blot techniques (EITB) for studying the specificities of antigens and antibodies separated by gel electrophoresis. *Methods Enzymol* 92: 377-391.
23. Garcia HH, Gilman RH, Martinez M, Tsang VCW, Pilcher JB, Herrera G, Diaz F, Alvarado M, Miranda E, 1993. Cysticercosis as a major cause of epilepsy in Peru. *Lancet* 341: 197-200.
24. Barkovich A, Citrin C, Klara P, Wippold F, Kattah J, 1986. Magnetic resonance imaging of cysticercosis. *West J Med* 145: 687-690.
25. Bryan RT, 1992. Current issues in cysticercosis: proteins, proglottids, pigs, and privies. Walker DH, ed. *Global Infectious Diseases: Prevention, Control and Eradication*. Vienna: Springer-Verlag, 181-203.
26. Wilson M, Bryan R, Fried J, Ware D, Schantz P, Pilcher JB, Tsang VCW, 1991. Clinical evaluation of the cysticercosis enzyme-linked immunoelectrotransfer blot in patients with neurocysticercosis. *J Infect Dis* 164: 1007-1009.
27. Heimans JJ, De Visser M, Polman CH, Nauta J, Kamphorst W, Troost D, 1990. Accuracy and interobserver variation in the interpretation of computed tomography in solitary brain lesions. *Arch Neurol* 47: 520-523.